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Preface

Germicidal effects of UV-C were demonstrated both in air and water in the 1st half of the 20th century. UV-C was first used in a drinking water plant in Marseille in 1910. By the 1930's the germicidal action curve was known and UV-C upper air was used in a school to prevent infection by measles. In the 40's UV-C upper air was being used in health care settings to disinfect the air. In the 50's/60's it was used in air disinfection for TB prevention and next to drinking water disinfection it was more and more used in several other applications such as in food &beverage preparation /industrial processes and residential water to disinfect water from private wells. By 1980 there were more than 1000 drinking water plants using UV-C disinfection in Europe. In the US waste water plants started using UV-C instead of chlorine. In the 90's UV-C started to be used on cooling coils in HVAC installations. The growth of the UV-C disinfection market continued after 2000 in existing applications expanding globally and in various new applications such as aquaria, fish ponds, swimming pools, air handlers, surface disinfection etc.

Whilst UV-C can be used as the exclusive solution in some applications, it is often used in tandem with other techniques. It follows that a single technology solution approach is unlikely to be ideal. It also follows that since UV-C is so simple and energy effective, it is perhaps wise to consider this option first.

Signify has been closely associated with progress in this field by developing, manufacturing and marketing lamps generating UV-C and continues to research new lamp configurations.



1. Micro-organisms

General

A micro-organism is a living organism that is too small to see with the naked eye. Bacteria, yeasts, fungi, viruses, and single-celled parasites are micro-organisms. Micro-organisms are all around us. In the air, in (natural) water, in our food and in and on animals.

We live in balance with micro-organisms. When a micro-organism can no longer reproduce and grow we call it cellular death or in practical terms: loss of the ability for cell division.

To kill/remove micro-organisms, cleaning and disinfection is needed. By cleaning we mean the removal of dirt and other unwanted material. In addition, the micro-organisms present in the dirt are removed. Subsequently, on indication, disinfection may be necessary. This reduces the number of living micro-organisms to an acceptable level in order to prevent the spread of micro-organisms.

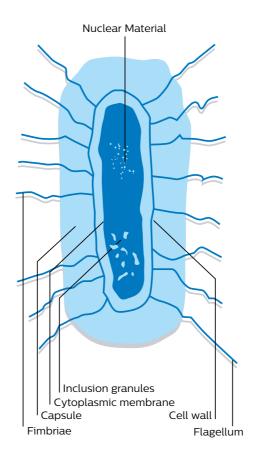


Figure 1. The main components of a typical bacterial cell.

1.1 Bacteria and spores

1.1.1 Bacteria

Bacteria do not have a cell nucleus and are single-celled. Bacteria can multiply through cell division.

Bacteria are very small and cannot be seen with the naked eye. They are everywhere and most bacteria are not harmful, in fact they are indispensable. Without bacteria, humans could not live. Bacteria convert food and help separate waste. This process takes place in the gastrointestinal tract and is called digestion. In addition, on our skin, we have skin bacteria that provide a natural defense against anything we get on our hand, for example. There are however bacteria that can harm human health and cause severe infections like E.coli, Streptococcus, Staphylococcus.



Figure 2. Some examples of bacteria varieties

1.1.2 Bacterial spores

Some bacteria can form spores. These spores can survive in unfavorable conditions. Once conditions are favorable again, the spore can germinate and grow into a normal cell and can then cause disease or spoilage.

1.2 Fungi and yeasts

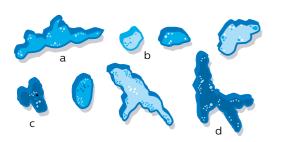


Figure 3. Brewer's yeast (Saccharomyces cerevisiae) in various stages of development: a. Various forms b. Yeast cell with spores c. Yeast spores d. Yeast spores after germination.

1.2.1 Fungi

Fungi, unlike bacteria, have a cell nucleus. They require nutrition to grow. Fungi are aerobic; they need oxygen to stay alive and grow. Fungi have filaments that branch and grow out. Spores are formed in these threads or at the tip. Under favourable conditions, a new fungus can form from this spore.

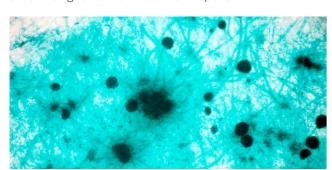


Figure 4. Mould culture.

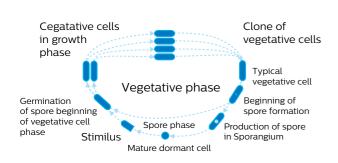


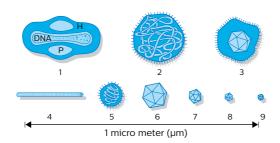
Figure 5. 'Life cycle' of spore formers.

1.2.2 Yeasts

Yeasts are single-celled fungi. Yeasts convert sugar into alcohol and carbon dioxide. Yeasts can be found in beer, wine, but also in bread. In addition to their usefulness, yeasts can also cause damage by spoiling food, but they can also cause infection.

13 Viruses

A virus is an incredibly small organism that can only multiply in living cells. The other cell does not survive this. So they live at the expense of the other cell. Viruses always cause illnesses, such as the flu or colds. Viruses can easily adapt, this is called mutating. If a virus mutates, it can cause the treatment that was normally effective to no longer be effective. Well known viruses are SARS-CoV-2, rotavirus, seasonal flu virus.



4. Tobacco mosaic virus

6. Insect polyhedral virus

5. Influenza virus

7. Adeno virus

8. Polyema virus

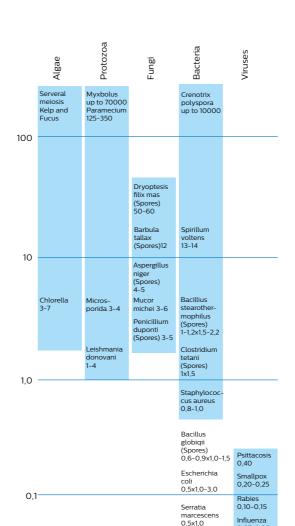
Figure 6. Relative shapes and sizes of some types of viruses.

- 1. Smallpox virus Abbreviations:
- DNA = virus DNA

- 3 Hernes virus
- P = elliptical protein body
- H = enveloping layers
- 2. Mumps virus
 - 9. Poliomyelitis virus



Figure 7. SARS-CoV-2 virus.



Influenza 0,07-0,08

Mosaic virus 0,042

Poliomey-litis 0,025-

Foot and 0,008-0,012

| nsions in µm 1 µm=0,001 | Dimensions in μm | 0,001 |
|-------------------------|------------------|-------|
| nsions in µm I µn | Dimensions in µm | |

Figure 8. Relative sizes of different types of micro-organisms.

1.4 Protozoa

Protozoa is an informal name for a group of unicellular organisms that live independently or parasitize other life forms. A well-known example of a protozoa is plasmodium, causative agent of malaria. Self-contained species of protozoa feed on organic material from certain types of micro-organisms. Protozoa are not classified as animals, but the name "protozoa" refers to the numerous, diverse species of single-celled organisms in which each cell contains a nucleus, which can move independently.

1.5 Algae

Algae is the informal name for a large collection of organisms that take their energy from sunlight producing Oxygen in the meantime. Algaes all contain the green pigment called chlorofyl for fotosyntheses and do not have a sterile (cells not participating in reproduction) layer of cells around their gametes (reproductive cell). Most are aquatic and autotrophic and lack many of the distinct cell and tissue types which are found in land plants.

2. Ultraviolet radiation

General

Ultraviolet (UV) is that part of electromagnetic light bounded by the lower wavelength extreme of the visible spectrum and the X-ray radiation band. The spectral range of UV light is, by definition between 100 and 400 nm (1 nm=10⁻⁹m) and is invisible to human eyes. Using the CIE classification the UV spectrum is subdivided into three bands:

UV-A (long-wave) from 315 to 400 nm UV-B (medium-wave) from 280 to 315 nm UV-C (short-wave) from 100 to 280 nm

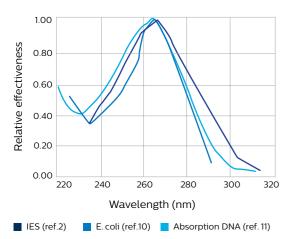


Figure 9. Germicidal action spectrum.

A strong germicidal effect is provided by the Light in the short-wave UV-C band (figure 9) In addition erythema (reddening of the skin) and conjunctivitis (inflammation of the mucous membranes of the eye) can also be caused by this form of Light. Because of this, when UV-C lamps are used, it is important to design systems to exclude UV-C leakage and so avoid these effects.

Self evidently people should avoid exposure to UV-C. Fortunately this is relatively simple, because it is absorbed by most materials, and even standard flat glass absorbs all UV-C. Exceptions are quartz and PTFE. Again fortuitously, UV-C is mostly absorbed by dead skin, so erythema can be limited. In addition UV-C does not penetrate to the eye's lens; nevertheless, conjunctivitis can occur and though temporary, it is extremely painful; the same is true of erythemal effects.

| Permissible UV-C Exposures | |
|------------------------------|----------------------|
| Duration of exposure per day | Irradiance (µW/cm²) |
| 8 hours | 0.2 |
| 4 hours | 0.4 |
| 2 hours | 0.8 |
| 1 hour | 1.7 |
| 30 minutes | 3.3 |
| 15 minutes | 6.6 |
| 10 minutes | 10 |
| 5 minutes | 20 |
| 1 minute | 100 |
| | |

Table 1. Permissible 254 nm UV exposures, according to ACGIH.

Where exposure to UV-C Light occurs, care should be taken not to exceed the threshold level norm. Figure 10 shows these values for most of the CIE UV spectrum. In practical terms, table I gives the American Congress of Governmental and Industrial Hygienist's (ACGIH) UV Threshold Limit Effective Irradiance Values for human exposure related to time. At this time it is worth noting that radiation wavelengths below 240 nm forms ozone, O3 from oxygen in air. Ozone is toxic and highly reactive; hence precautions have to be taken to avoid exposure to humans and certain materials.

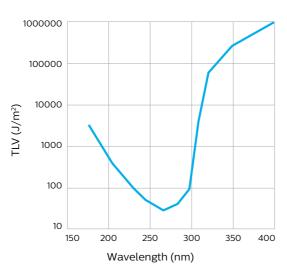


Figure 10. American Conference of Governmental Industrial Hygienists. (Accessed 2013, May 23). Ultraviolet Radiation: TLV® Physical Agents 7th Edition Documentation

2.1 Generation and characteristics of short-wave UV light

The most efficient source for generating UV-C is the low-pressure mercury discharge lamp, where on average 35% of input watts is converted to UV-C watts. The radiation is generated almost exclusively at 254 nm viz. at 85% of the maximum germicidal effect (figure 9). Philips' low pressure ultraviolet (TUV) lamps have an envelope of special glass that filters out ozone-forming radiation, in this case the 185 nm mercury line. The spectral transmission of this glass is shown in figure 11 and the spectral power distribution of these TUV lamps is given in figure 12.

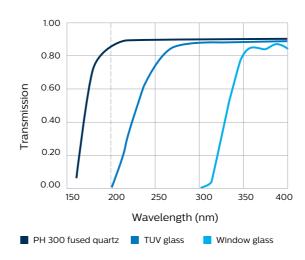


Figure 11. Special transmission of glasses (1 mm).

For various Philips low pressure mercury UV-C lamps the electrical and mechanical properties are identical to their lighting equivalents. This allows them to be operated in the same way i.e. using an electronic or magnetic ballast/starter circuit. For low pressure mercury lamps there is a relationship between lamp operating temperature and output. In low pressure lamps the resonance line at 254 nm is strongest at a certain mercury vapour pressure in the discharge tube. This pressure is determined by the operating temperature and optimises at a tube wall temperature of 40°C, corresponding with an ambient temperature of about 25°C. (See page 29, figure 26). It should also be recognised that lamp output is affected by air currents (forced or natural) across the lamp, the so called chill factor. The reader should note that, for some lamps, increasing the air flow and/or decreasing the temperature can increase the UV-C output. This is met in high output (HO) lamps viz. lamps with higher wattage than normal for their linear dimension. (See page 30, figure 28).

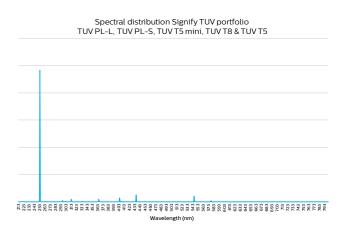


Figure 12. Spectral distribution Signify TUV portfolio.

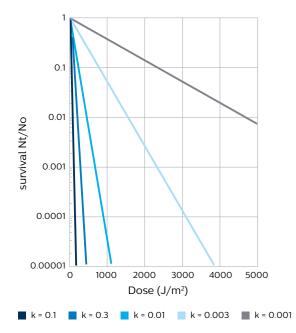


Figure 13. Survival of micro-organisms depending on dose and rate constant k.

2.2 Germicidal action

The UV light emitted by a source is expressed in watts (W) and the irradiation density is expressed in watts per square meter (W/m^2). For germicidal action dose is important. The dose is the irradiation density multiplied by the time (t) in seconds and expressed in joules per square meter (J/m^2). (1 joule is 1W.second).

From figure 10 it can be seen that germicidal action is maximised at 265 nm with reductions on either side. Low pressure lamps have their main emission at 254 nm where the action on DNA is 85% of the peak value and 80% on the IES curve. For wavelengths below 235 nm the germicidal action is not specified, but it is reasonable to assume that it follows the DNA absorption curve.

Micro-organisms effective resistance to UV light varies considerably. Moreover, the environment of the particular micro-organism greatly influences the radiation dose needed for its destruction. Water, for instance, may absorb a part of the effective radiation depending on the concentration of contaminants in it. Iron salts in solution ware well known inhibitors. Iron ions absorb the UV light. The survival of micro-organisms when exposed to UV light is given by the approximation:

- · In is the natural logarithm
- N, is the number of germs at time t
- No is the number of germs before exposure
- · k is a rate constant depending on the species
- E_{eff} is the effective irradiance in W/m² The product E_{eff} t is called the effective dose

Heff and is expressed in W.s/m² of J/m²

It follows that for 90% kill equation 2 becomes

$$2.303 = kH_{eff}$$

Some k value indications are given in table 2, where they can be seen to vary from $0.2 \text{ m}^2/\text{J}$ viruses and bacteria, to 2.10^{-3} for mould spores and 8.10^{-4} for algae. Using the equations above, figure 13 showing survivals or kill % versus dose, can be generated.

| Bacteria Dose Bacillus anthracis 45.2 B. megatherium sp. (spores) 27.3 B. megatherium sp. (veg.) 13.0 B. parathyphosus 32.0 B. suptilis 71.0 B. suptilis spores 120.0 Campylobacter jejuni 11.0 Clostridium tetani 120.0 Corynebacterium diphteriae 33.7 Dysentery bacilli 22.0 Eberthella typhosa 21.4 Escherichia coli 30.0 Klebsiella terrifani 26.0 Legionella pneumophila 9.0 Micrococcus candidus 60.5 Micrococcus sphaeroides 100.0 Mycobacterium tuberculosis 60.0 Neisseria catarrhalis 44.0 Phytomonas tumefaciens 44.0 Pseudomonas aeruginosa 55.0 Pseudomonas fluorescens 35.0 Proteus vulgaris 26.4 Salmonella enteritidis 40.0 Salmonella paratyphi 32.0 Salmonella typhimurium 80.0 Sarcina lutea 197.0 Sarcina lutea 197.0 Staphylococcus albus 18.4 Staphylococcus albus 18.4 Staphylococcus lacus 61.5 Streptococcus lactus 61.5 Streptococcus viridans 20.0 | |
|--|-------|
| B. megatherium sp. (spores) B. megatherium sp. (veg.) B. megatherium sp. (veg.) B. megatherium sp. (veg.) B. suptilis B. suptilis B. suptilis spores B. suptilis spor | |
| B. megatherium sp. (veg.) B. parathyphosus 32.0 B. suptilis 71.0 B. suptilis spores 120.0 Campylobacter jejuni 11.0 Clostridium tetani 120.0 Corynebacterium diphteriae 33.7 Dysentery bacilli 22.0 Eberthella typhosa Escherichia coli 30.0 Klebsiella terrifani 26.0 Legionella pneumophila Micrococcus candidus Micrococcus sphaeroides 100.0 Mycobacterium tuberculosis 60.5 Mycobacterium tuberculosis 44.0 Phytomonas tumefaciens Pseudomonas aeruginosa Pseudomonas fluorescens 35.0 Proteus vulgaris Salmonella enteritidis 40.0 Salmonella paratyphi 32.0 Salmonella paratyphi 32.0 Salmonella typhimurium 80.0 Sarcina lutea 197.0 Seratia marcescens 24.2 Shigella paradysenteriae 16.3 Shigella sonnei Shigella sonnei Streptococcus hemoluticus Streptococcus lactus 61.5 | 0.051 |
| B. parathyphosus 32.0 B. suptilis 71.0 B. suptilis spores 120.0 Campylobacter jejuni 11.0 Clostridium tetani 120.0 Corynebacterium diphteriae 33.7 Dysentery bacilli 22.0 Eberthella typhosa 21.4 Escherichia coli 30.0 Klebsiella terrifani 26.0 Legionella pneumophila 9.0 Micrococcus candidus 60.5 Micrococcus sphaeroides 100.0 Mycobacterium tuberculosis 60.0 Neisseria catarrhalis 44.0 Phytomonas tumefaciens 44.0 Pseudomonas aeruginosa 55.0 Proteus vulgaris 26.4 Salmonella enteritidis 40.0 Salmonella paratyphi 32.0 Salmonella typhimurium 80.0 Seratia marcescens 24.2 Shigella paradysenteriae 16.3 Shigella sonnei 30.0 Streptococcus lacus 61.5 Streptococcus lactus 61.5 Streptococcus lactus 61.5 Streptococcus lactus 61.5 | 0.084 |
| B. suptilis pores 120.0 Campylobacter jejuni 11.0 Clostridium tetani 120.0 Corynebacterium diphteriae 33.7 Dysentery bacilli 22.0 Eberthella typhosa 21.4 Escherichia coli 30.0 Klebsiella terrifani 26.0 Legionella pneumophila 9.0 Micrococcus candidus 60.5 Micrococcus sphaeroides 100.0 Mycobacterium tuberculosis 60.0 Neisseria catarrhalis 44.0 Phytomonas tumefaciens 44.0 Pseudomonas aeruginosa 55.0 Pseudomonas fluorescens 35.0 Proteus vulgaris 26.4 Salmonella enteritidis 40.0 Salmonella paratyphi 32.0 Salmonella typhimurium 80.0 Sarcina lutea 197.0 Seratia marcescens 24.2 Shigella paradysenteriae 16.3 Shigella sonnei 30.0 Streptococcus lactus 61.5 Streptococcus hemoluticus 21.6 Streptococcus lactus 61.5 | 0.178 |
| B. suptilis spores Campylobacter jejuni Clostridium tetani 120.0 Corynebacterium diphteriae 33.7 Dysentery bacilli 22.0 Eberthella typhosa 21.4 Escherichia coli 30.0 Klebsiella terrifani 26.0 Legionella pneumophila 9.0 Micrococcus candidus 60.5 Micrococcus sphaeroides 100.0 Mycobacterium tuberculosis 60.0 Neisseria catarrhalis 44.0 Phytomonas tumefaciens Pseudomonas aeruginosa Pseudomonas fluorescens 35.0 Proteus vulgaris Salmonella enteritidis 40.0 Sarcina lutea Sarcina lutea Seratia marcescens 24.2 Shigella paradysenteriae Shigella sonnei Streptococcus hemoluticus Streptococcus lactus 11.0 120.0 | 0.072 |
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| Corynebacterium diphteriae 33.7 Dysentery bacilli 22.0 Eberthella typhosa 21.4 Escherichia coli 30.0 Klebsiella terrifani 26.0 Legionella pneumophila 9.0 Micrococcus candidus 60.5 Micrococcus sphaeroides 100.0 Mycobacterium tuberculosis 60.0 Neisseria catarrhalis 44.0 Phytomonas tumefaciens 44.0 Pseudomonas aeruginosa Pseudomonas fluorescens 35.0 Proteus vulgaris 26.4 Salmonella enteritidis 40.0 Salmonella paratyphi 32.0 Salmonella typhimurium 80.0 Sarcina lutea 197.0 Seratia marcescens 24.2 Shigella paradysenteriae 16.3 Shigella sonnei 30.0 Streptococcus albus Streptococcus hemoluticus Streptococcus lactus 61.5 | 0.209 |
| Dysentery bacilli 22.0 Eberthella typhosa 21.4 Escherichia coli 30.0 Klebsiella terrifani 26.0 Legionella pneumophila 9.0 Micrococcus candidus 60.5 Micrococcus sphaeroides 100.0 Mycobacterium tuberculosis 60.0 Neisseria catarrhalis 44.0 Phytomonas tumefaciens 44.0 Pseudomonas aeruginosa 55.0 Pseudomonas fluorescens 35.0 Proteus vulgaris 26.4 Salmonella enteritidis 40.0 Salmonella paratyphi 32.0 Salmonella typhimurium 80.0 Seratia marcescens 24.2 Shigella paradysenteriae 16.3 Shigella sonnei 30.0 Streptococcus albus 18.4 Streptococcus hemoluticus 21.6 Streptococcus lactus 61.5 | 0.019 |
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| Escherichia coli Xlebsiella terrifani 26.0 Legionella pneumophila 9.0 Micrococcus candidus 60.5 Micrococcus sphaeroides 100.0 Mycobacterium tuberculosis 60.0 Neisseria catarrhalis 44.0 Phytomonas tumefaciens 44.0 Pseudomonas aeruginosa 55.0 Proteus vulgaris 26.4 Salmonella paratyphi 32.0 Salmonella paratyphi 32.0 Salmonella typhimurium 80.0 Sarcina lutea 197.0 Seratia marcescens 24.2 Shigella paradysenteriae 16.3 Shigella sonnei 30.0 Streptococcus aureus 26.0 Streptococcus hemoluticus Streptococcus lactus 61.5 | 0.105 |
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| Legionella pneumophila 9.0 Micrococcus candidus 60.5 Micrococcus sphaeroides 100.0 Mycobacterium tuberculosis 60.0 Neisseria catarrhalis 44.0 Phytomonas tumefaciens 44.0 Pseudomonas aeruginosa 55.0 Pseudomonas fluorescens 35.0 Proteus vulgaris 26.4 Salmonella enteritidis 40.0 Salmonella paratyphi 32.0 Salmonella typhimurium 80.0 Sarcina lutea 197.0 Seratia marcescens 24.2 Shigella paradysenteriae 16.3 Shigella sonnei 30.0 Spirillum rubrum 44.0 Staphylococcus albus 18.4 Staphylococcus faecalis 44.0 Streptococcus hemoluticus 21.6 Streptococcus lactus 61.5 | 0.077 |
| Micrococcus candidus Micrococcus sphaeroides 100.0 Mycobacterium tuberculosis 60.0 Neisseria catarrhalis 44.0 Phytomonas tumefaciens 44.0 Pseudomonas aeruginosa 55.0 Pseudomonas fluorescens 35.0 Proteus vulgaris 26.4 Salmonella enteritidis 40.0 Salmonella paratyphi 32.0 Salmonella typhimurium 80.0 Sarcina lutea 197.0 Seratia marcescens 24.2 Shigella paradysenteriae 16.3 Shigella sonnei 30.0 Spirillum rubrum 44.0 Staphylococcus albus 18.4 Staphylococcus faecalis 44.0 Streptococcus hemoluticus 21.6 Streptococcus lactus 61.5 | 0.089 |
| Micrococcus sphaeroides Mycobacterium tuberculosis 60.0 Neisseria catarrhalis 44.0 Phytomonas tumefaciens 44.0 Pseudomonas aeruginosa 55.0 Proteus vulgaris 26.4 Salmonella enteritidis 40.0 Salmonella paratyphi 32.0 Salmonella typhimurium 80.0 Seratia marcescens 24.2 Shigella paradysenteriae 16.3 Shigella sonnei 30.0 Spirillum rubrum 44.0 Staphylococcus albus Streptococcus faecalis Streptococcus lactus 61.5 | 0.256 |
| Mycobacterium tuberculosis Neisseria catarrhalis 44.0 Phytomonas tumefaciens 44.0 Pseudomonas aeruginosa 55.0 Pseudomonas fluorescens 35.0 Proteus vulgaris 26.4 Salmonella enteritidis 40.0 Salmonella paratyphi 32.0 Salmonella typhimurium 80.0 Sarcina lutea 197.0 Seratia marcescens 24.2 Shigella paradysenteriae 16.3 Shigella sonnei 30.0 Spirillum rubrum 44.0 Staphylococcus albus Streptococcus faecalis 44.0 Streptococcus hemoluticus 21.6 Streptococcus lactus 61.5 | 0.038 |
| Neisseria catarrhalis Phytomonas tumefaciens 44.0 Pseudomonas aeruginosa Pseudomonas fluorescens 35.0 Proteus vulgaris Salmonella enteritidis 40.0 Salmonella paratyphi 32.0 Salmonella typhimurium 80.0 Sarcina lutea 197.0 Seratia marcescens 24.2 Shigella paradysenteriae 16.3 Shigella sonnei 30.0 Spirillum rubrum 44.0 Staphylococcus albus Staphylococcus faecalis 44.0 Streptococcus hemoluticus Streptococcus lactus 61.5 | 0.023 |
| Phytomonas tumefaciens Pseudomonas aeruginosa Pseudomonas fluorescens Proteus vulgaris Salmonella enteritidis Salmonella paratyphi Salmonella typhimurium Sarcina lutea Seratia marcescens Shigella paradysenteriae Shigella sonnei Staphylococcus albus Streptococcus faecalis Streptococcus lactus 44.0 44.0 55.0 56.4 46.4 40.0 58.0 | 0.038 |
| Pseudomonas aeruginosa 55.0 Pseudomonas fluorescens 35.0 Proteus vulgaris 26.4 Salmonella enteritidis 40.0 Salmonella paratyphi 32.0 Salmonella typhimurium 80.0 Sarcina lutea 197.0 Seratia marcescens 24.2 Shigella paradysenteriae 16.3 Shigella sonnei 30.0 Spirillum rubrum 44.0 Staphylococcus albus 18.4 Staphylococcus aureus 26.0 Streptococcus hemoluticus 21.6 Streptococcus lactus 61.5 | 0.053 |
| Pseudomonas fluorescens 35.0 Proteus vulgaris 26.4 Salmonella enteritidis 40.0 Salmonella paratyphi 32.0 Salmonella typhimurium 80.0 Sarcina lutea 197.0 Seratia marcescens 24.2 Shigella paradysenteriae 16.3 Shigella sonnei 30.0 Spirillum rubrum 44.0 Staphylococcus albus Streptococcus faecalis 44.0 Streptococcus hemoluticus 21.6 Streptococcus lactus 61.5 | 0.053 |
| Proteus vulgaris 26.4 Salmonella enteritidis 40.0 Salmonella paratyphi 32.0 Salmonella typhimurium 80.0 Sarcina lutea 197.0 Seratia marcescens 24.2 Shigella paradysenteriae 16.3 Shigella sonnei 30.0 Spirillum rubrum 44.0 Staphylococcus albus 18.4 Staphylococcus faecalis 44.0 Streptococcus hemoluticus 21.6 Streptococcus lactus 61.5 | 0.042 |
| Salmonella enteritidis Salmonella paratyphi 32.0 Salmonella typhimurium 80.0 Sarcina lutea 197.0 Seratia marcescens 24.2 Shigella paradysenteriae 16.3 Shigella sonnei 30.0 Spirillum rubrum 44.0 Staphylococcus albus Staphylococcus aureus 26.0 Streptococcus faecalis 44.0 Streptococcus hemoluticus 21.6 Streptococcus lactus 61.5 | 0.065 |
| Salmonella paratyphi 32.0 Salmonella typhimurium 80.0 Sarcina lutea 197.0 Seratia marcescens 24.2 Shigella paradysenteriae 16.3 Shigella sonnei 30.0 Spirillum rubrum 44.0 Staphylococcus albus 18.4 Staphylococcus aureus 26.0 Streptococcus faecalis 44.0 Streptococcus hemoluticus 21.6 Streptococcus lactus 61.5 | 0.086 |
| Salmonella typhimurium 80.0 Sarcina lutea 197.0 Seratia marcescens 24.2 Shigella paradysenteriae 16.3 Shigella sonnei 30.0 Spirillum rubrum 44.0 Staphylococcus albus 18.4 Staphylococcus aureus 26.0 Streptococcus faecalis 44.0 Streptococcus hemoluticus 21.6 Streptococcus lactus 61.5 | 0.058 |
| Sarcina lutea 197.0 Seratia marcescens 24.2 Shigella paradysenteriae 16.3 Shigella sonnei 30.0 Spirillum rubrum 44.0 Staphylococcus albus 18.4 Staphylococcus aureus 26.0 Streptococcus faecalis 44.0 Streptococcus hemoluticus 21.6 Streptococcus lactus 61.5 | 0.072 |
| Seratia marcescens 24.2 Shigella paradysenteriae 16.3 Shigella sonnei 30.0 Spirillum rubrum 44.0 Staphylococcus albus 18.4 Staphylococcus aureus 26.0 Streptococcus faecalis 44.0 Streptococcus hemoluticus 21.6 Streptococcus lactus 61.5 | 0.029 |
| Shigella paradysenteriae 16.3 Shigella sonnei 30.0 Spirillum rubrum 44.0 Staphylococcus albus 18.4 Staphylococcus aureus 26.0 Streptococcus faecalis 44.0 Streptococcus hemoluticus 21.6 Streptococcus lactus 61.5 | 0.012 |
| Shigella sonnei 30.0 Spirillum rubrum 44.0 Staphylococcus albus 18.4 Staphylococcus aureus 26.0 Streptococcus faecalis 44.0 Streptococcus hemoluticus 21.6 Streptococcus lactus 61.5 | 0.095 |
| Spirillum rubrum 44.0 Staphylococcus albus 18.4 Staphylococcus aureus 26.0 Streptococcus faecalis 44.0 Streptococcus hemoluticus 21.6 Streptococcus lactus 61.5 | 0.141 |
| Staphylococcus albus 18.4 Staphylococcus aureus 26.0 Streptococcus faecalis 44.0 Streptococcus hemoluticus 21.6 Streptococcus lactus 61.5 | 0.077 |
| Staphylococcus aureus 26.0 Streptococcus faecalis 44.0 Streptococcus hemoluticus 21.6 Streptococcus lactus 61.5 | 0.053 |
| Streptococcus faecalis 44.0 Streptococcus hemoluticus 21.6 Streptococcus lactus 61.5 | 0.126 |
| Streptococcus hemoluticus 21.6 Streptococcus lactus 61.5 | 0.086 |
| Streptococcus lactus 61.5 | 0.052 |
| · | 0.106 |
| Streptococcus viridans 20.0 | 0.037 |
| | 0.115 |
| Sentertidis 40.0 | 0.057 |
| Vibrio chlolerae (V.comma) 35.0 | 0.066 |
| Yersinia enterocolitica 11.0 | 0.209 |

Table 2. Doses for 10% survival under 254 nm radiation (J/m^2) and rate constant k (m^2/J), Ref 2, 3, 4, 5, 6 and 7.

Note: this is a guideline. Sensitivity data may vary between

| Dose | k |
|------|----------------------|
| 39 | 0.060 |
| 33 | 0.070 |
| 60 | 0.038 |
| 60 | 0.038 |
| 60 | 0.038 |
| 80 | 0.019 |
| | 39 33 60 60 |

| Mould spores | | |
|------------------------|------|--------|
| Aspergillus flavus | 600 | 0.003 |
| Aspergillus glaucus | 440 | 0.004 |
| Aspergillus niger | 1320 | 0.0014 |
| Mucor racemosus A | 170 | 0.013 |
| Mucor racemosus B | 170 | 0.013 |
| Oospora lactis | 50 | 0.046 |
| Penicillium digitatum | 440 | 0.004 |
| Penicillium expansum | 130 | 0.018 |
| Penicillium roqueforti | 130 | 0.018 |
| Rhizopus nigricans | 1110 | 0.002 |

| Virus | | |
|-----------------|-----|-------|
| Hepatitis A | 73 | 0.032 |
| Influenza virus | 36 | 0.064 |
| MS-2 Coliphase | 186 | 0.012 |
| Polio virus | 58 | 0.040 |
| Rotavirus | 81 | 0.028 |
| SARS-CoV-2 | 18 | 0.13 |

| Protozoa | | |
|------------------------|----|-------|
| Cryptosporidium parvum | 25 | 0.092 |
| Giardia lamblia | 11 | 0.209 |
| Giardia lamblia | 11 | 0.209 |

| Algae | | |
|--------------------|------|--------|
| Blue Green | 3000 | 0.0008 |
| Thlorella vulgaris | 120 | 0.019 |



General

In practice, disinfection applications and design factors are governed by three main factors:

A. The effective dose (Heff)

Effective dose is the product of time and effective irradiance (the irradiance that makes a germicidal contribution). However, dose is severely limited by its ability to penetrate a medium. Penetration is controlled by the absorption co-efficient; for solids total absorption takes place in the surface; for water, depending on the purity, several 10s of cm or as little as a few microns can be penetrated before 90% absorption takes place.

B. Design considerations

UV-C radiation can produce conjunctivitis and erythema, therefore people should not be exposed to it at levels more than the maximum exposure given in figure 9. It follows that this needs to be taken into consideration when designing purification equipments. UV-C lamps can be applied and are used for all three states of matter, viz. gases (air), liquids (mainly water) and solids (surfaces) with greatest technical success in those applications where the absorption coefficient is smallest.

However, some notable success has been achieved in applications where, despite a disadvantageous absorption, "thin film" or closed circuit (recycling the product) design techniques have provided effective solutions.

C. Lamps

Philips ranges of lamps are available for purification purposes:

- · Classic Philips T5 and T8 TUV lamps
- High output Philips TUV lamps
- Philips PL-S and PL-L twin-tube compact TUV lamps
- Philips extreme power technology (XPT) amalgam lamps in various diameters

All of these are based on low pressure mercury technology. Increasing the lamp current of low pressure lamps produces higher outputs for lamps of the same length; but at the cost of UV efficiency (UV watts/input watts); this is due to higher selfabsorption levels, and temperature influences. The application of mercury amalgams, rather than pure mercury, in the lamps corrects for the latter.

The choice of the lamp type depends on the specific application. In most cases the low pressure types are the most attractive. This is because UV-C lamps are highly efficient in destroying micro-organisms, hence there is limited need for high wattage lamps. For water purification, low and medium pressure are both used, although the choice is not necessarily based on UV-C efficacy. Initial total systems costs, including metalwork and space limitations, can be the driving factor rather than efficacy.

D. Systems

Near lamps Signify provides also inhouse manufactured electronic lamp drivers to offer a complete system solution for ultimate performance.

3.1 Air purification (Ref. 12, 13)

Good results are obtained with this form of purification because air has a low absorption coefficient and hence allows UV-C to attack micro-organisms present. In addition, two other beneficial conditions are generally present, viz. random movements allowing bacteria etc. to provide favorable molecular orientations for attack and high chances of "closed circuit" conditions, that is second, third and more recycle opportunities. From this, it is evident that air purification is an important application for UV light.

Even in the simplest system (natural circulation) there is an appreciable reduction in the number of airborne organisms in a room. Thus the danger of airborne infection, a factor in many illnesses, is considerably reduced.

Presently, there are five basic methods of air purification using UV lamps viz:

- a. Ceiling or wall mounted Philips TUV lamps.
- b. Philips TUV lamps (in upwards-facing reflectors) for upper-air irradiation.
- c. Philips TUV lamps (in downwards-facing reflectors) for irradiation of the floor zone (often in combination with b.).
- d. Philips TUV lamps in air ducts sometimes in combination with special dust filters.
- e. Philips TUV lamps, incorporated in stand-alone air cleaners with a simple filter.

3.1.1 Ceiling-mounted Philips TUV lamps

This method is used in those cases where either the interior is unoccupied or where it is possible for the occupants to take protective measures against UV-C light. These protective measures entail covering the:

Face

Glass spectacles, closefitting goggles or plastic face visors

Hande

Gloves (for long exposure, special plastic is preferable to rubber)

Head and neck

Head cover

Note: Normal glasses and plastics can be used to give protection, because they transmit little or no UV-C; some exceptions are special UV glasses, quartz and certain PTFEs.

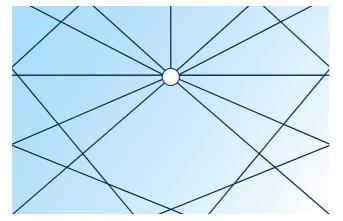
3.1.2 Philips TUV lamps for upper-air irradiation using upward facing reflectors

This method of purification can be used to combat bacteria and moulds; it also has the advantage that it can be used in occupied interiors without the occupants using protective clothing. The lamps should be mounted in suitable reflectors and aimed to emit no radiation below the horizontal.

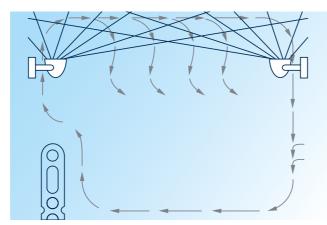
The reflectors should be mounted more than 2.10 m above the floor, the lower air thus entirely free of any direct UV light. Air above the 2.10 m level maintains a low germ level, because it subject to direct UV-C light.

Free convection of air without forced ventilation causes air movements of about 1.5 – 8 m³ per minute, thus producing exchanges between the upper treated and lower untreated parts of the room. The process reduces air contamination to fractions of that before the TUV lamps were activated. As an indication for general applications in a simple room, or enclosure, it is advisable to install an effective UV-C level of 15–20 mW total fixture output per m³ total room volume* or 30–50 microwatt per cm² in the irradiated zone.**

Figure 14. Various principles of air purifications.



a. Ceiling mounted lamps



b. Upwards facing reflectors



c. Horizontal beam with louvred luminaire

* Mphaphlele, Dharmadhikari, Jensen, et al.: Trial of Upper Room Ultraviolet Air Disinfection

3.1.3 Philips TUV lamps for irradiation of the floor zone using downward facing reflectors

This method is for use in those cases where it is important that the entire room air, even at floor level is rendered as sanitary as possible. In this case, lamps supplementing those irradiating the upper air should be fitted in downward-aimed reflectors at about 60 cm above the floor

In methods 3.1.1, 3.1.2 and 3.1.3 person detectors/systems can be used to deactivate TUV lamps, if necessary.

3.1.4 Philips TUV lamps in air ducts

In this method, all the conditioned air is subjected to radiation prior to entry in the room. The injected air can be disinfected to a specified killing level, depending upon the number of lamps installed and the dwell time, that is the time spent in the effective killing region of the lamp(s); by definition this takes the dimensions of the air duct into consideration. Such systems have a controlled flow rate and their performance can be predicted theoretically. Certain aspects should be borne in mind, however

- These installations are most suitable for bacteria or viruses; most moulds have higher resistances to UV, so the air flow rate is not likely to allow a sufficient dwell time to produce a high enough effective dose.
- Dust filters should be installed to prevent the lamps from becoming soiled and hence seriously reducing their effective emission.
- The number of lamps required in an air purifying chamber in an air duct system is dependent on the required degree of disinfection, the airflow rate, the ambient temperature, the humidity of the air and the UV-C-reflecting properties of the chamber walls.

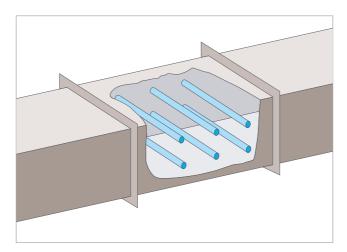


Figure 15. Basic arrangement of Philips TUV lamps in an air duct for room purification.

The advantage of purifying air prior to it entering a room is that there is then no limit to the maximum permitted radiation dose, since humans are totally shielded.

Designing duct systems needs to account for practical issues, such as large temperature and humidity variations caused by exterior weather variations, if only because air is often drawn from outside, then released into a room after a single pass over the lamps. Recycling part of the air will allow multiple passes, hence improving system efficiency.

Lining the UV lamps section with aluminum, also increases efficiency. The lamps and the wall of the duct should be easily accessible to permit regular cleaning and easy maintenance, another reason for a modular design. Micro-organisms exposed to UV, experience a normal exponential decrease in population, as already expressed on page 10:

$N_{t}/N_{o} = \exp(-kE_{off}t)$

The rate constant defines the sensitivity of a microorganism to UV light and is unique to each microbial species. Few airborne rate constants are known with absolute certainty. In water based systems, Escherichia coli are often used as test organism. It is however not an airborne pathogen. For aerosolization tests, often the innocuous Serratia marcescens is used.

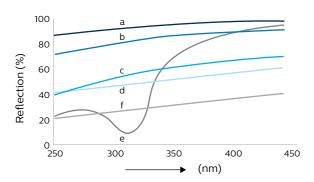
Points to remember when constructing Philips TUV lamp installations in air ducts:

- The surface of the chamber walls should have a high reflectance to UV 254 nm, for example by using anodised aluminum sheet (reflectance 60–90 per cent).
- The lamps should be so arranged that there are no 'shadow' areas.
- Lamps should be mounted perpendicular to the direction of the airflow.
- Use HO lamps having better performance in moving air.
- Lamps should be changed after the nominal lifetime; an elapsed time meter will help.
- An external pilot light should be used to indicate that the lamps are functioning.

^{**} DHHS (NIOSH) Publication No. 2009-105

Reflectance of various materials to UV 254 nm

The graphs shown give the spectral reflectance of various metals (figure 16) and organic substances (figure 17) to radiation of different wavelengths. These graphs demonstrate the importance of determining a material's 254 nm reflectance. As can be seen, high reflectance to visible radiation is not consistent with high reflectance to short-wave UV light.



d. Nickel

Figure 16. Metal surfaces (indicative numbers).

- a. Aluminum foil
- e. Silver c. Evaporated aluminum f. Stainless steel

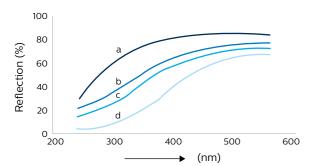


Figure 17. Organic substances.

- a Bleached cotton c Linen d White wool b. White paper
- Materials with a high reflectance to 254 nm are used to construct reflectors for both direct and upper-air irradiation. Material with a low reflectance to 254 nm are used where UV light has to be absorbed after performing its function. This latter is necessary to avoid the consequences resulting from the unwanted 254 nm reflections, so ceilings and walls should be treated with a low reflectance material.

3.1.5 Philips TUV lamps in stand-alone units

Recently this method has gained commercial favor by meeting a growing need for a better Indoor Air Quality, (IAQ). Closed stand- alone devices are safe, simple and flexible. In essence the units consist of Philips TUV lamps, mostly PL-L types driven by high frequency ballasts, mounted inside a "light trap" container. The unit incorporates a fan that firstly draws air across a filter, then across the lamp(s). Single and multiple lamp options can be built into a small outer using either single or double-ended lamp options.

For maximum design flexibility. PL-L and PL-S lamps offer the best solutions, because their dimensions are compact, so reducing unit size and because their single ended configuration allows more mounting options.

The units have the benefits of portability and hence more mounting positions viz. wall, floor or ceiling mounted in either permanent or temporary options. A feature of their design is that cleaning and lamp and filter replacement is easy. Additionally their portability can be used to produce immediate results. Variation in UV-C dose can be achieved both by varying the number of lamps and their wattage (see also dimming below). As an example, it is possible to use the same physical design dimensions for PL-L lamps with a nominal wattage range between 18 and 95W HO, in single and multi lamp variants. Commercial products are known for as few as 1 x PL-L 18W and as many as 4 x PL-L 95W HO lamps inside the same container, giving a unit capable of producing a 25-fold difference in effective dose. PL-L lamps are more flexible; they have readily available and competitively priced electronic regulating (dimming) ballasts to vary UV output in a simple reliable fashion. Ballasts can be single, double and in the case of 18 W, four lamp versions. This adds to the flexibility of portable units.

| Material | Reflectance % |
|-----------------------------|---------------|
| Aluminum: untreated surface | 40-60 |
| treated surface | 60-89 |
| sputtered on glass | 75-85 |
| 'ALZAK' - treated aluminum | 65-75 |
| 'DURALUMIN' | 16 |
| Stainless steel/Tin plate | 25-30 |
| Chromium plating | 39 |
| Various white oil paints | 3-10 |
| Various white water paints | 10-35 |
| Aluminum paint | 40-75 |
| Zinc oxide paint | 4-5 |
| Black enamel | 5 |
| White baked enamel | 5-10 |
| White plastering | 40-60 |
| New plaster | 55-60 |
| Magnesium oxide | 75-88 |
| Calcium carbonate | 70-80 |
| Linen | 17 |
| Bleached wool | 4 |
| Bleached cotton | 30 |
| Wallpapers:ivory | 31 |
| white | 21-31 |
| red printed | 31 |
| ivory printed | 26 |
| brown printed | 18 |
| White notepaper | 25 |

Table 3. Reflectance of various materials to UV-254 nm radiation.

3.2 Surface disinfection

Surface disinfection generally requires high-intensity short-wave UV light. Mostly this means TUV lamps are mounted close to the surface requiring to be kept free from biofilm or to be disinfected.

The success of surface disinfection depends largely on the surface irregularity of the material to be disinfected, because UV light can only inactivate those micro-organisms that it hits with a sufficient dose. Thus disinfection can only be successful if the entire surface is exposed to UV light. Micro-organisms sitting in "holes" in a surface are not likely be overcome by reflections from the hole walls, as can be deduced from the reflectances shown in table 3.

In practice, solid surfaces, granular material and packaging (whether plastic, glass, metal, cardboard, foil, etc.) are disinfected or maintained germ-free by means of intensive, direct irradiation. Additionally, disinfected material can be kept largely germ-free throughout its further processing by irradiating the air along its path.



Figure 18. UV-C disinfection robot.

3.3 Water and other liquids disinfection

UV-C radiation is capable of penetrating liquids with varying degrees of efficiency. From a treatment view, liquids can be regarded as similar to air so the further the UV light is able to penetrate the liquid, the more efficient is its action. The degree of efficiency thus greatly depends on the liquid and more particularly its absorption coefficient at 254 nm (table 4). As an example, natural water's transparency to 254 nm may vary by as much as a factor of 10 or more from place to place. Polluted industrial water often needs filtration followed by disinfection; here UV-C is growing with many thousands of systems in use in North America and Europe, each with a multitude of lamps. Often UV light may supplement or replace conventional chlorination measures (see later). UV-C has advantages over chlorinating techniques, because it produces far fewer noxious by-products and it is unaffected by the pH of the water or its temperature. The reader should note that the latter comment refers to the radiation, not to the lamp, or its environment as described earlier.

Micro-organisms are far more difficult to kill in humid air, or in a liquid environment, than in dry air. This is because they limit transmission of 254 nm radiation. In more quantitative terms liquids decrease the UV-C intensity exponentially according to the formula

 $E_x = E_0.e^{-\alpha(x)}$ Ex intensity at depth x E0 incident intensity α absorption coefficient

Liquids with a high α can only be disinfected when they are exposed as thin films. A rough indication to estimate penetration depth is $1/\alpha$, at this depth the irradiation level will have fallen to 1/e or to 37%. To overcome wall effects where liquids are notoriously static, turbulence or rigorous stirring is necessary for better disinfection, agitation helps orientate microorganisms hidden behind particles.

Iron salts (as well as other inorganic salts) and suspended matter in liquids will decrease the effectiveness of UV-C radiation.

Additionally, it is feasible that organic compounds, in particular, those susceptible to bond fissure under UV light, can change the texture and taste of the liquid being treated.

Hence experimentation is needed. In round terms the effective depth of penetration for a 90% kill may thus vary from 3 m for distilled water, down to 12 cm for normal drinking water and even less in wines and syrups (2.5 mm), see table 4.

The penetration depths cause more special techniques to be applied to allow 254 nm radiation to penetrate sufficiently, these include generating "thin films" and or slow speed presentation to the radiation, so that a sufficient dose can be applied.

If an UV lamp has to be immersed in a liquid, it should be enclosed in a quartz or UV-C transparent PTFE sleeve.

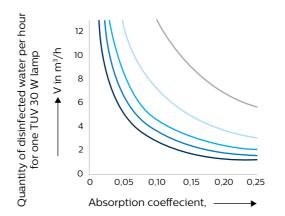


Figure 19. Volume of disinfected water \boldsymbol{V} as a function of the absorption coefficient.

 α (for distilled water α = 0.007–0.01/cm, for drinking water α = 0.02–0.1/cm) with respect to different degrees of purification (in terms of Escherichia coli).

| Example of absorption coeffici | ents |
|--------------------------------|------------|
| Liquid | |
| Wine, red | 30 |
| Wine, white | 10 |
| Beer | 10-20 |
| Syrup, clear | 2-5 |
| Syrup, dark | 20-50 |
| Milk | 300 |
| Distilled water | 0.007-0.01 |
| Drinking water | 0.02-0.1 |

Table 4. Absorption coefficient (α) of various liquids to UV-254 nm per cm depth.



4. Applications

General

The main application areas for UV-C lamps may be briefly classified below, although there are many other areas, where the lamps may be employed for various purposes.

- Water purification
- Municipal drinking water
- Municipal waste water
- · Residential drinking water
- Water coolers dispensers
- Semiconductors process water
- Spas and swimming pools
- Cooling towers
- · Fish ponds and aquariums
- Air purification
- Cooling coils

4.1 Water purification (Ref. 7, 14)

A wide variety of micro-organisms in the water can cause disease, especially for young and senior people, who may have weaker immune systems. UV light provides purification without the addition of chemicals that can produce harmful by-products and add unpleasant taste to water. Additional benefits include easy installation, low maintenance and minimal space requirements.

UV has the ability to inactivate bacteria, viruses and protozoa. Each type of organism requires a specific dose for inactivation. Viruses require higher doses than bacteria and protozoa. Understanding the organisms to be neutralised will help to determine the size of the UV system that will be required. For example, to kill 99,9% of E.coli, a UV dose of 90 J/m² or 9 mW.sec/cm² is required.

UV installations are suitable for industrial, municipal and residential markets

The quality of the water has an important effect on the performance of UV systems. The common factors that have to be considered are the total concentration of suspended solids and the UV transmittance. Various organic and inorganic compounds can absorb UV.

When there is uncertainty about what may be present in the water, the UV transmittance should be tested. Most drinking water supplies have UV transmittances between 85% and 95%.

Separate treatment technologies often are required to improve the water quality before purification:

- Sediment filters or particle filters, to remove particles that "shadow" microbes or absorb UV.
- Carbon filters or reverse osmosis, which remove organic compounds.
- · Chemical methods to remove contaminants such as iron

UV is often used in conjunction with Reverse Osmosis (RO) applications. Purification prior to the RO systems increases the durability of the RO membrane by reducing the accumulation of bacterial biofilms.

The reactor of a UV purification device must be designed to ensure that all microbes receive sufficient exposure of the UV.

Most manufacturers of UV equipment use low pressure mercury lamps. High output, (HO) versions and amalgam lamps are rapidly becoming popular. High capacity drinking water and waste water systems feature medium pressure mercury technology.

The temperature of the lamp surface is one of the most critical factors for UV reactor design. The UV efficiency of the the low pressure mercury lamp (UV output per consumed electrical wattage) strongly depends on the bulb temperature. (See page 29, figure 26).

The diameter of the protective quartz sleeve should be carefully adapted to the specific power of the lamp (Watts per unit of arc length), as well as temperature and velocity of the water flow.

As the lamp ages, the UV output declines due to solarization of the lamp (glass or quartz) envelope. The quoted dose for a specific unit is the minimum dose that will be delivered at the end of the lamp's life. Most manufacturers offer electronic power supplies, that are more efficient (up to 10%) and operate at lower temperatures. Such power supplies normally withstand wide fluctuations in supply voltage, still providing a consistent current to the lamps.

Factors, that should be considered, when, choosing the right size of UV equipment, in order, to achieve the desired purification objectives are peak flow rate, the required dose and the UV transmittance of the water.

Theoretical calculations should be validated by bioassay tests, for a variety of conditions that include flow rates and variable water quality.

4.1.1 Municipal waste water

Chlorine has been used to disinfect waste water for over a century. However, while chlorine is very effective, it is also associated with environmental problems and health effects. Chlorination byproducts in waste water effluents are toxic to aquatic organisms, living in surface waters. Chlorine gas is hazardous to human beings. UV irradiance has proven to be an environmentally responsible, convenient and cost-effective way to disinfect public waste water discharges. UV purification is much safer than waste water systems that rely on chlorine gas, as it eliminates transport and handling of large quantities of this hazardous chemical. More than thousands of waste water installations all over the world rely on UV purification these days. The required UV dose levels depend on the upstream processes, and range, taking into account flow rates and UV transmittance of the water, between 50 and 100 m J/cm².



Figure 20. Waste water system.

4.1.2 Municipal drinking water

Purification of drinking water by UV light is a wellestablished technology in Europe. Hundreds of European public water suppliers have by now incorporated UV disinfection. The driving force in Europe was to inactivate bacteria and viruses, but avoid use of chlorine. Recent studies regarding potential negative health effects of purification by-products have led to a critical view on chlorine. A few fatal waterborne outbreaks of cryptosporidiosis in North America have proven the fact that existing purification and filtration technologies could not guarantee to eliminate cryptosporidium oocysts from the water.

Cryptosporidium parvum is a human pathogen, capable of causing diarrhoeal infections, sometimes even leading to death. The organism can be shed as an environmentally resistant form (oocyst) and persists for months.



Figure 21. UV drinking water plant.

Cryptosporidium is almost completely resistant against chlorine. Ozone can be effective, but the water quality and temperature play a significant role. Its small size makes it difficult to remove by standard filter techniques.

Studies have verified that UV can achieve significant inactivation of cryptosporidium at very modest doses. Exposures as low as 10 mJ/cm² will result in a more than 4- log reduction of concentration.

The effectiveness of UV for cryptosporidium removal, together with stricter limits on purification by-products have paved the way for UV disinfection in North America. Due to their high UV efficiency, low pressure HO lamps have found their way in many municipal UV drinking water facilities. However, as space always will be a problem, the high intensity medium pressure lamps will be favorite, especially when existing drinking water plants have to be upgraded with a UV extension.

4.1.3 Residential drinking water

Classic Point of Use (POU) / or Point-of-Entry (POE) UV disinfection systems consist of a low-pressure mercury UV lamp, protected against the water by a quartz sleeve, centered into a stainless steel reactor vessel.

The UV output is monitored by an appropriate UV sensor, providing visual or audible indicators of the UV lamp status. To improve taste and odor of the water POU systems are often used in conjunction with an active carbon filter.

The ANSI/NSF Standard 55 (UV Microbiological Water Treatment Systems) establishes the minimum requirements a manufacturer will need to become certified for a Class A or B UV system.

Class A POU/POE devices are designed to disinfect micro-organisms, including bacteria and viruses, from contaminated water to a safe level. Waste water is specifically excluded from being used as feedwater. The UV system has to produce a UV dose of 40 mJ/cm².

Class A devices are required to have a UV sensor, alarming when the proper dose is not reaching the water.

Class B POU systems are designed for supplemental bacterial treatment of treated and disinfected public drinking water. Such devices are not intended for disinfection of microbiologically unsafe water. The systems are capable of delivering a UV dose of at least 16 mJ/cm² at 70% of the normal UV output or alarm set point.

4.1.4 Water coolers, dispensers

Water vending machines store and dispense water that is non-chlorinated. The machines must be licensed by local health service departments. One of the requirements for the license is that the vending machine is equipped with a purification unit to reduce the number of bacteria and other micro-organisms.

Bottled water coolers, which also dispense nonchlorinated water, are not required to contain a purification unit. However, without an active purification system, also bottled water cooler reservoirs are subject to biofilm growth. Such biofilms act like a breeding place for bacteria, protected by the gel-like substance. Bacteria contamination, regardless of whether it is non-harmful or even beneficial, is not a quality to be associated with drinking water. To avoid biofilm growth often simple UV reactors are being introduced.



Figure 22. POU residential drinking water UV Purification device.

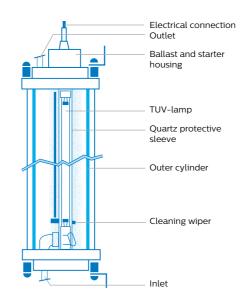


Figure 23. Basic sketch of TUV lamp operated water-purifying unit for general use.

4.1.5 Semiconductors process water

Organic compounds, present in the rinse water, can affect production yields and product quality in the semiconductor industry. The total organic carbon (TOC) contamination level is specified to be less than one part per billion (ppb) for ultrapure water, used for this application. UV light represents a powerful technology that has been successfully introduced in the production of ultrapure water for semiconductor, pharmaceutical, cosmetics and healthcare industries.

Its powerful energies can be applied, not only for purification, but also TOC reduction and destruction of ozone and chlorine.

Two different UV wavelengths are employed, 254 nm and 185 nm. The 254 nm energy is used for disinfection. It can also destroy residual ozone, present in the water. The 185 nm radiation decomposes the organic molecules. It carries more energy than the 254 nm and is able to generate hydroxyl free radicals from water molecules. These hydroxyl radicals are responsible for oxidizing the organics to carbon dioxide and water molecules. 185 nm radiating lamps are made of special quartz, with high transmittance for the lower wavelengths. Typical dosage requirements range from 100 to 500 mJ/cm². Philips XPT amalgam lamps in a 185 nm version can provide excellent solutions.

4.1.6 Spas and swimming pools

Philips TUV lamps are used to supplement the traditional methods of water treatment. Importantly, with UV-C as a supplement, less chlorine is needed for the same result. This is welcome both for those with allergies and those with a distaste for chlorine. The reason that UV-C is not suitable for sole use is that swimming pool water circulation has to take into consideration solids, inorganic compounds, hence filtration and chemical processes are also needed.

A standard technique is to circulate part of the water through a continuous flow UV-C device, thus creating a partial closed loop system; this in tandem with the chlorinator produces effective disinfection. It can lower the chlorine dose up to 50%.



Figure 24. Water purification system for a private swimming pool.

4.1.7 Miscellaneous

Fish ponds

Fish pond owners are often troubled by phototrophic micro-organisms. These are typical water organisms widely distributed in both fresh and salt water. Phototrophic bacteria contain photosynthetic pigment and hence they are strongly colored and appear as dense suspensions of either green, olive, purpleviolet, red, salmon or brown. Seasonal effects may lead to massive growth ('flowering of the water') as light helps chlorophyll synthesis.

If algae are to be destroyed or their growth inhibited, either a high dose of UV 254 nm radiation is needed or a long irradiation time. These conditions can be met relatively easily by creating a closed loop system whereby the water is presented to the UV-C source a number of times per day. The lamp is encased in a quartz tube. In practice, it has been found that, for instance, a Philips TUV PL-S 5W lamp in series with a filter can keep a 4.5 K liter (1,000 UK gallons) pond clear. For larger pond or pool volumes higher output lamps are needed on a pro rata scale. The process is thought to be that algae are split, recombine to form larger molecular chains, which can be removed by the filter, or are so large that they sink to the bottom of the pond.

Cooling towers

Cooling towers and re-circulating loops are often dirty, warm and rich in bio-nutrients. They are perfect breeding places for micro-organisms.

Chemical compounds, like chlorine or ozone, are fed into the system in regular intervals, to control the rate of biological growth. UV will substantially decrease the costs of purification, without any safety or environmental issues.

Aquariums

Aquariums present two problems: one is that they become swamped with algae; the second is that parasites may cause fish diseases. Both can occur in either freshwater or marine aquariums; warm water provides an excellent condition for micro-organisms

and the lighting features used also promotes algae growth. The same system as used for ponds is advocated, using no more than a Philips TUV PL-S 5W lamp for a private aquarium. A low pump speed will create a long dwell time across the lamp, so helping both bacteria kill rate and algae agglomeration. Using UV-C in ponds and aquariums is also beneficial because it can destroy parasites introduced by new fish; the latter can be catastrophic in many cases. UV-C treatment provides an effective solution particularly for suspended zoospores. Multiplication does not take place and aquariums can be free of parasites within a very short time. Even affected fish soon cease to display symptoms of morbidity.

Other applications using ultraviolet (UV) for water purification are: fish farming, ballast water for ships, agriculture, etc.







4.2 Air purification

Indoor air is trapped, often re-circulated and always full of contaminants such as bacteria, viruses, moulds, mildew, pollen, smoke and toxic gasses from building materials. Increasing levels of such contaminants act as triggering mechanisms for a variation of diseases of which asthma is the most prominent.

For offices and in industrial environments, so called HEPA (High Efficiency Particulate Air) filters can be installed in HVAC ductwork. Very fine fibers, pressed together, form a structure with openings, too small for most particulate contaminants. Such filters are effective, but always will give rise to considerable drop in air pressure. In recent days, growing concern for indoor air quality has lead to new measures. Application of UV in air ducts for ventilation, heating and cooling purposes has proven to provide adequate protection against airborne pathogens.

For domestic use some very different basic types can be considered:

- Fiber mesh filters, generally designed for a particle size of 25 microns or larger.
- Activated carbon filters, which will neutralise some gasses, smoke and odors.
- Electronic air cleaners, which charge particles such as dust, pollen and hair. The charged materials are attracted by a series of opposite polarity charged metal plates.
- Ozone and ion generators.
- UV light, the only treatment, truly lethal to micro-organisms.

With patients and visitors bringing in pathogens that can be transmitted by air, wards, clinics, waiting and operation rooms and similar areas should be protected against the risk of infection in personnel and patient populations, if possible at a reasonable cost!

Common traditional disease controlling methods in hospitals are:

- Ventilation: dilution of potentially contaminated air with uncontaminated air.
- · Negative pressure isolation rooms.
- HEPA (High Efficiency Particulate Air) filtration.

UV-C irradiation provides a potent, cost effective solution to upgrade protection against infection. (Ref. 12, 13)

Especially, upper-air purification has proven to be very effective to supplement existing controls for TBC and other airborne diseases (Ref. 8). Many disease-causing organisms circulate on air currents in "droplet nuclei", 1 to 5 micron in size, that are expelled with a cough, sneeze or even with speech. These droplet nuclei can be inhaled, spreading infections. It is estimated that up to 99% of airborne pathogens are destroyed with adequate air circulation and UV exposure.



4.3 Surface disinfection

On surfaces biofilm can grow and surfaces can be contaminated with micro-organism which can cause infections, diseases

4.3.1 Cooling coils

Air conditioner cooling coils are almost always wet and dusty and thus can serve as an ideal breeding ground for moulds, that can produce allergens.

Coil irradiation with UV drastically reduces the growth of moulds. At the same time heat exchange efficiency is improved and pressure drops decrease. As the coils are constantly irradiated, only a modest UV irradiance is required.

4.3.2 Disinfection units

UV-C disinfection chambers are used for disinfection of items like cell phones, handhelds and other everyday objects by destroying the bacteria's ability to multiply and spread disease.

4.3.3 Food industry

In the food industry, UV-C can disinfect transport belts and packaging materials (yoghurt cups, drinking bottles etc.). It can also be used to directly treat products to increase shelf life and preserve nutritional value (fewer preservatives required).

4.3.4 Treatment room in healthcare settings

In hospitals and elderly homes, there is a high risk of surface contamination with multidrug resistant pathogens causing hospital acquired illnesses. UV-C is used to decontaminate surfaces by fixed installations or robots.

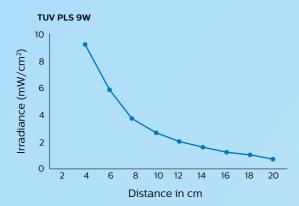
4.3.5 Miscellaneous

In high contact areas such as schools, retail outlets, industry, offices and public transportation, UV-C can be used for a deep disinfection of surfaces (e.g., floors, walls, desks).

5. Lamp data

General

For a complete survey, see separate product data brochures.



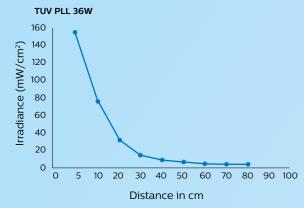
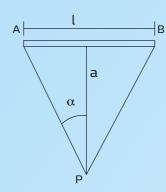


Figure 24 and 25. Demonstrate the variation of UV irradiance with the distance to the lamps.

5.1 UV irradiance values

The irradiance E on a small surface in point P on a distance a from an ideal linear radiation source AB of length 1 amounts to:



$$E = \frac{\varphi}{2\pi^2 \cdot l \cdot a} (2\alpha + \sin \alpha)$$

E= irradiance in W/m²

 φ = UV-C output of the lamp in W

 π = the number Pi

l = distance between the lamp electrodes in m

a = distance from the lamp to point P in m

 α = angle to the lamp electrode in rad

 φ is the total radiation flux (in W). This formula is taken from: H. Keitz, Light calculations and measurements, Philips Technical Library, MacMillan and Co Ltd, 1971.

For a large distance to the lamp we get:

$$E = \frac{\varphi}{\pi^2.a^2}$$
.....(2)

At shorter distances the irradiance is proportional to

$$E = \frac{\varphi}{2\pi I a}$$
 (a < 0.5 I)....(3)

For a variety of low pressure mercury TUV lamps, the irradiance values at 1 meter distance are expressed below.

| Irradiance at 1m distance* | |
|----------------------------------|--------|
| Lamptype | μW/cm² |
| Philips TUV 4W T5 | 10 |
| Philips TUV 6W T5 | 17 |
| Philips TUV 8W T5 | 24 |
| Philips TUV 11W T5 | 30 |
| Philips TUV 16W T5 | 44 |
| Philips TUV 20W T5 | 68 |
| Philips TUV 25W T5 | 75 |
| Philips TUV 10W T5 (T8 adapters) | 28 |
| Philips TUV 15W T8 | 50 |
| Philips TUV F17T8 (18W) | 60 |
| Philips TUV 25W T8 | 75 |
| Philips TUV 30W T8 | 110 |
| Philips TUV 36W T8 | 132 |
| Philips TUV 55W HO | 175 |
| Philips TUV 75W HO | 230 |
| Philips TUV PL-S 5W | 14 |
| Philips TUV PL-S 7W | 21 |
| Philips TUV PL-S 9W | 29 |
| Philips TUV PL-S 11W | 44 |
| Philips TUV PL-L 13W | 42 |
| Philips TUV PL-L 18W | 58 |
| Philips TUV PL-L 24W | 93 |
| Philips TUV PL-L 36W | 140 |
| Philips TUV PL-L 55W HF | 212 |
| Philips TUV PL-L 35W HO | 111 |
| Philips TUV PL-L 60W HO | 235 |
| Philips TUV PL-L 95W HO | 310 |
| Philips TUV 24T5 HE | 106 |
| Philips TUV 24T5 HO | 172 |
| Philips TUV 36T5 HE | 145 |
| Philips TUV 36T5 HO | 227 |
| Philips TUV 48T5 HE | 170 |
| Philips TUV 48T5 HO | 289 |
| Philips TUV 64T5 HE | 270 |
| Philips TUV 64T5 HO | 392 |
| Philips TUV 130W XPT | 465 |
| Philips TUV 180W XPT | 615 |
| Philips TUV 200W XPT | 663 |
| Philips TUV 325W XPT | 1132 |
| Philips TUV 330W XPT | 957 |
| Philips TUV 350W XPT | 1180 |
| Philips TUV 800W XHO | 2585 |
| Philips TUV 230W XPT | 159 |
| Philips TUV 260W DIM | 163 |
| Philips TUV 260W HO | 200 |
| Philips TUV 335W XPT | 189 |
| Philips TUV 335W HO | 250 |
| | |

Table 6. Irradiance values of Philips TUV lamps at a distance of 1 meter.

5.2 Influence of temperature

The UV efficiency of low-pressure mercury lamps is directly related to the (saturated) mercury pressure. This pressure depends on the lowest temperature spot on the lamp. Optimum UV efficiency is achieved when this temperature is approximately 40 °C, see figures 26 and 27. For low pressure mercury amalgam lamps the mercury pressure in the discharge is regulated by the amalgam spots that keep the mercury pressure constant over a temperature range between 80 and 120 degrees Celsius (fig. 27). Moving air has a strong impact on the tube wall temperature. The cooling effects of air streams (and lower ambient temperatures) can be compensated by over-powering the lamps. Figure 28 shows this effect, comparing standard Philips TUV PL-L 36W lamps with high output 60W versions, having the same dimensions. For low pressure mercury lamps lowest temperature spot to be measured is behind electrode or middle of the lamp, whichever is lower. For amalgam lamps this temperature is to be measured on the amalgam spot.

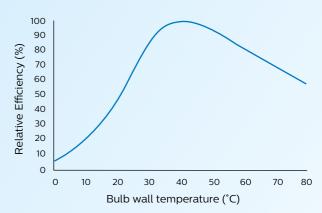


Figure 26. Temperature dependence of low pressure mercury lamps

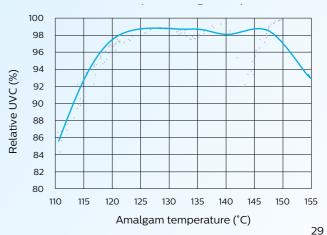


Fig 27. Temperature dependence of T6 amalgam lamps

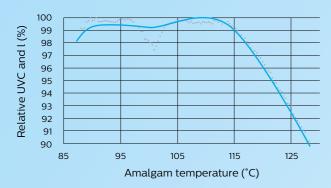


Fig 27. Temperature dependence of T8, T10, T12 amalgam lamps

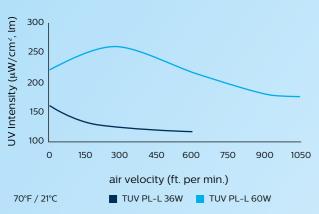


Figure 28. UV vs Windchill Factor.

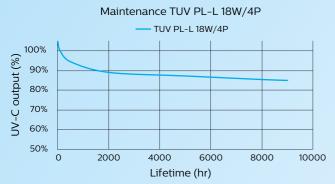


Figure 29. UV-C maintenance curve.

5.3 Lamp life

The life of low pressure mercury lamps (TUV) depends on:

- electrode geometry
- · lamp current
- · noble-gas filling
- switching frequency
- $\cdot \ \text{ambient temperature} \\$
- circuitry

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Photographs by courtesy of:

- Finsen Tech, UK (www.finsentech.com/products/thor-uvc) (Fig 18)
- Trojan Technologies, London Ontario, Canada (www.trojanuv.com) (Figures 20 and 21)
- Kent, India (www.kent.co.in) (Fig 22)
- VGE BV, the Netherlands (www.vgebv.nl) (Fig 24)

140 20 30 10 3 1 0,3 0,1 0,03 Number of switchings per 3 hours

Lamp and driver need to be perfectly matched to

operated and frequently switched. Even when the

reach the optimal lamp life both when continuously

drivers are meeting these requirements an increased

number of switches will negatively impact the lamp

life. This effect of number of switches on lamp life is

shown in figure 30 nominal lamp life is reached if the

lamps are switched 8 times in 24 burning hours. If the

number of switches increases to 24 times in 24 hours

burning time, the lamp life will decrease with 30%.

Figure 30. Lamp life.

180

5.4 No Ozone emission:

The Philips TUV lamp portfolio is produced in soft glass (TUV PL-L, TUV PL-S, TUV T5, TUV T8 and TUV TLmini) or Quartz glass (TUV XPT lamps), both glass types are blocked for emission below 200 nm and therefore the Philips TUV lamps do not emit any 185 nm. Ozone.



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